

Targeted Protein and Peptide Quantitation Using Agilent LC/MS and Skyline Software

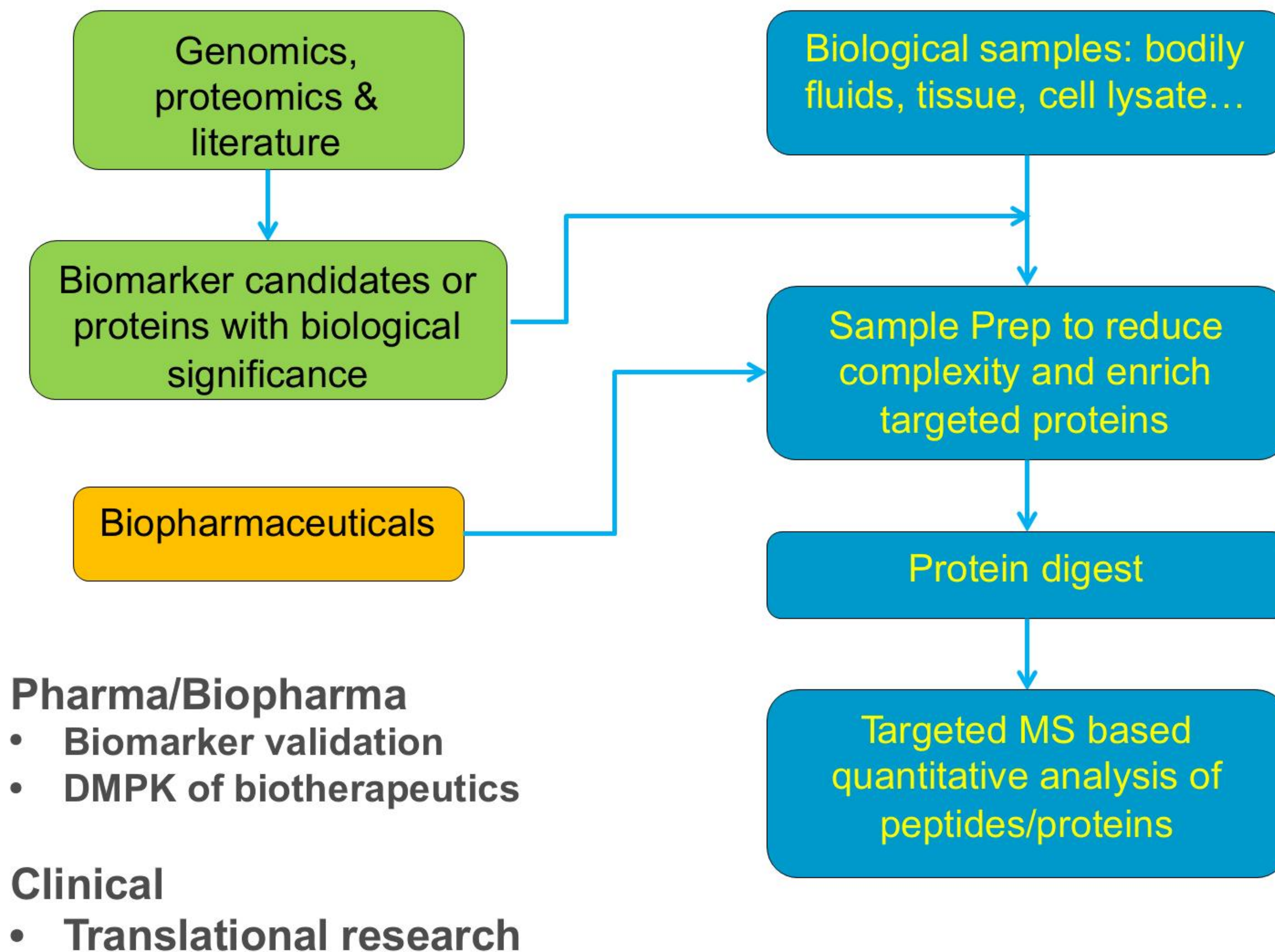


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Agilent Technologies

Targeted Protein Quantitation



MRM Based Quantitation Workflow Overview

- Selection of target proteins
- Selection of target peptides that represent the protein list
- Selection of best charge states and MRM transitions for each targeted peptide
- Optimization of collision energy for each MRM transition
- Application of the assays to the detection and quantitation of the proteins



MRM Assay Development – Current Approaches

1. Data dependent acquisition (Auto MS/MS on a QTOF) → Database search → Create MRM list or spectral library to use in Skyline
2. MS2 Scan followed by Product ion scans on a QQQ
3. Publically available databases (PeptideAtlas, the Global Proteome machine Database, etc.).
4. Skyline MRM predictions.



MRM Assay Development – Current Approaches From Skyline Prediction

MacCoss Lab Software

Home

Skyline

Skyline Targeted Proteomics Environment

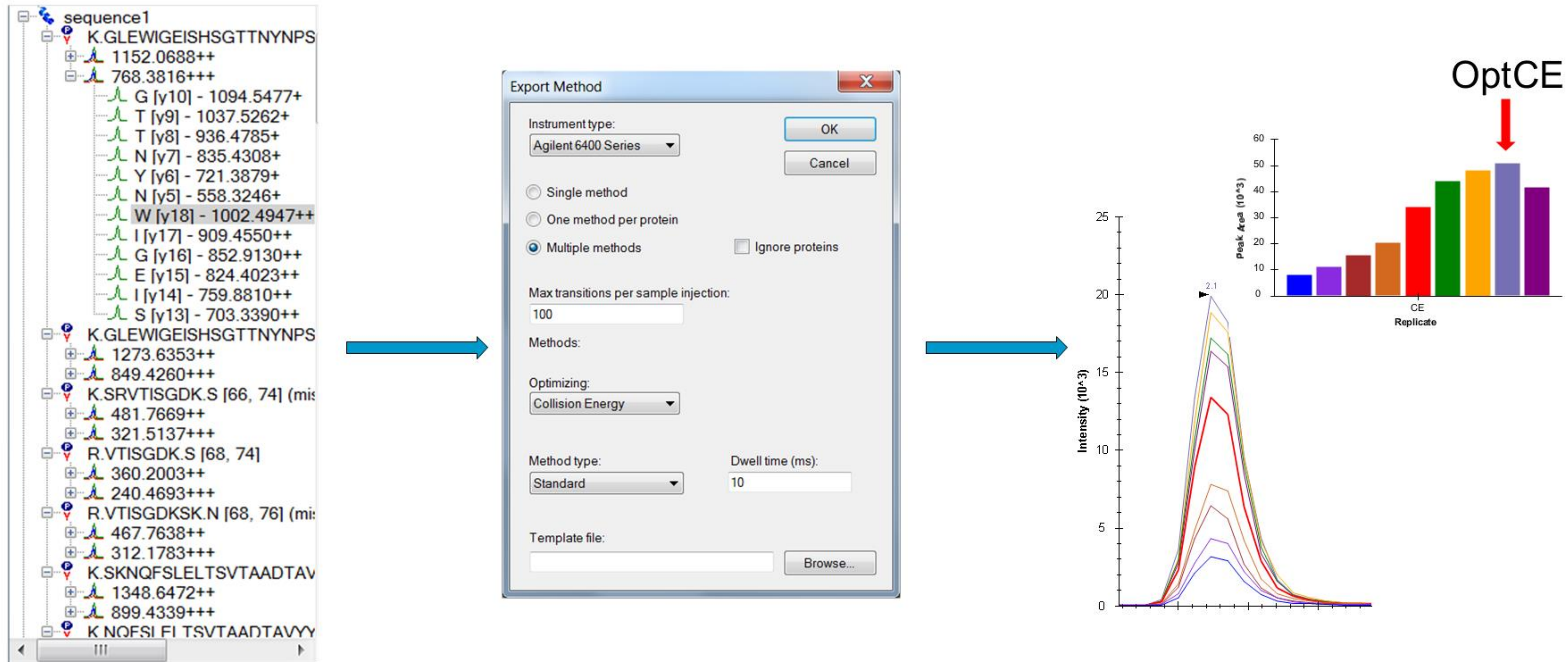
Join over 3500 other proteomics investigators:
Sign up now to get email notification of releases and events, and access to free support.
 (only if you are not already a registered user)

Download & Install
 Skyline 2.5 - 3
 Skyline 2.5 - 6
 VIDEO TRAILER
 External Tools

- Open Source software
- Multi-vendor software
- Rapidly evolved using feedback from top-labs
- Widely used and highly regarded



MRM Assay Development – Current Approaches From Skyline Prediction



For this mAb, the entire list of > 40 peptides need to be evaluated. The number can be even bigger if including peptides with miscleavages.

CE Optimization Using Agilent Automation Tool

The image displays two software windows. The top window is 'Skyline - HSA-Feb13-auto.sky', showing a 'Tools' menu with 'Agilent Automation' highlighted. The bottom window is 'Skyline Automation - D:\chris\HSA-Feb13-auto.sky', showing a project configuration interface with three steps: Step-A, Step-B, and Step-C. Step-A is currently selected, showing settings for 'Export method name' (Demo), 'Single method', 'Max transitions per sample injection' (200), 'Optimizing' (None), 'Method type' (Standard), and 'Dwell time (ms)' (5). The right side of the window shows a workflow diagram and three tables for sample data across the steps.

Automation tool automatically creates QQQ methods and acquire data for CE optimization, reloads and analyzes the results, export the final optimized QQQ method (MRM, dMRM, tMRM) and use it to runs any real samples queued up in the worklist

(Step-A)				
	Sample prefix	Standard	Start position	P1-A1
1	Standard1	P1-A1	Demo.m	Step-A_Demo1.d

(Step-B)				
	Sample prefix	Standard	Start position	P1-A1
1	Standard1	P1-A1	Demo_0001.m	Step-B_Demo_00011.d

(Step-C)				
	Sample prefix	Samples	Start position	P1-A1
1	Sample1	P1-A1	Demo.m	Step-C_Demo1.d
2	Sample2	P1-B1	Demo.m	WorklistData2.d
3	Sample3	P1-B2	Demo.m	WorklistData3.d
4	Sample4	P1-B3	Demo.m	WorklistData4.d
5	Sample5	P1-B4	Demo.m	WorklistData5.d

Project is created at: D:\MassHunter\Data\chris\demo.s. (3/4/2013 7:57:09 AM) Total samples: 12



Challenges

- No prediction for representative peptides/preferred charge state.
- Questionable prediction for modified peptides.
- Numerous runs for collision energy optimization based on the number of proteins and peptides.



Simplified MRM Method Development:

Agilent All Ions MS/MS on a QTOF + Skyline SW

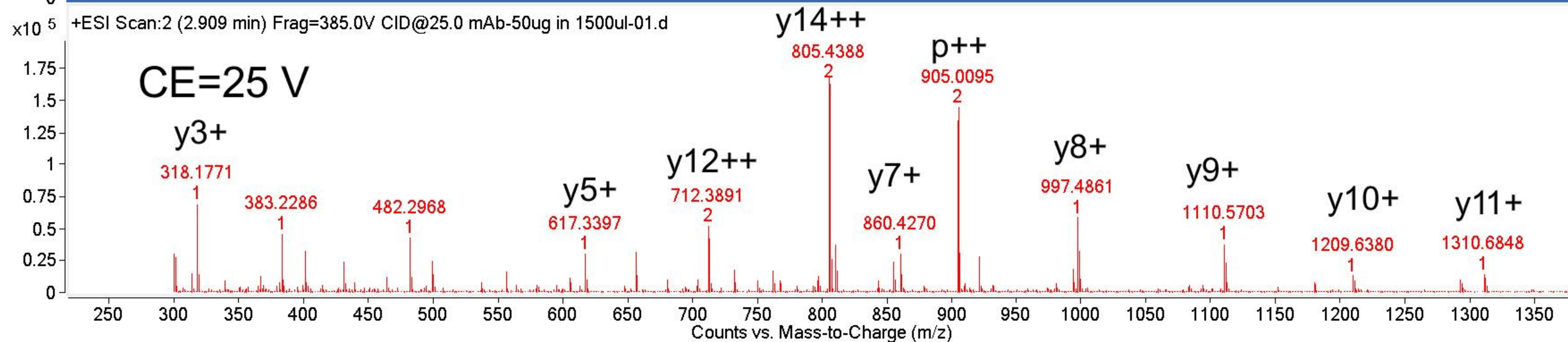
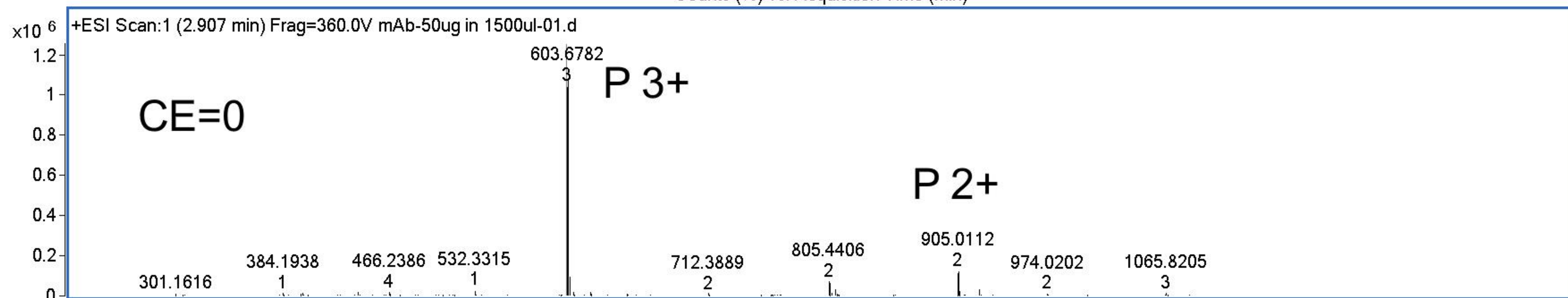
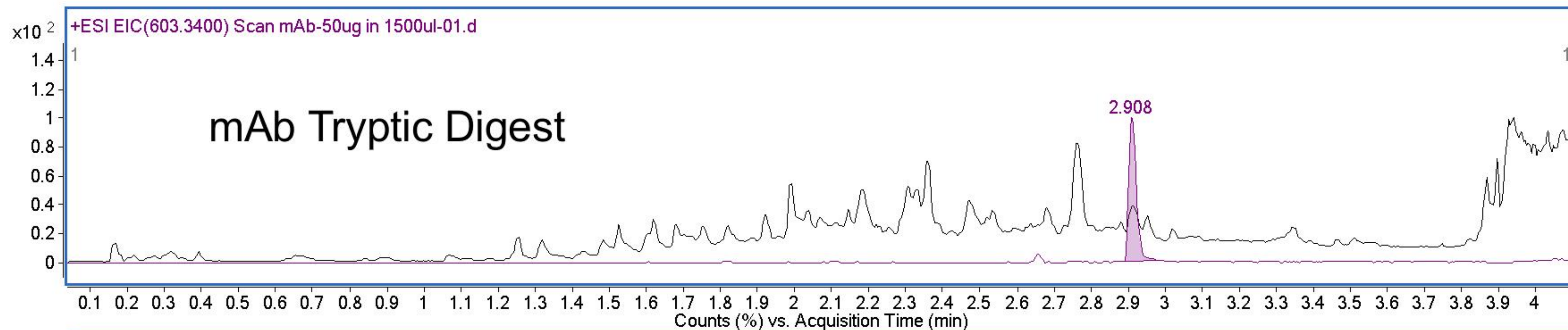
Agilent All Ions MS/MS :

- Data independent acquisition, no precursor selection, short cycle time
- Easy acquisition set up
- Stores MS and MS/MS spectra for all detectable peptides

The screenshot shows the 'Method Editor' window for a Q-TOF instrument. The 'Q-TOF' tab is selected, and the 'Acquisition' sub-tab is active. The method name is 'FK_AIM_4CE(0-10-20-40)_4Hz_5-98(11m).m'. The 'Ion Source' is set to 'Dual ESI', 'Ion Polarity' is 'Positive', 'Data Storage' is 'Both', and 'LC Stream' is 'MS'. The 'Stop Time' is set to 'No Limit/As Pump'. The 'Time Segment and Experiment #' table shows a segment from 0 to 0.5 minutes, with experiment 3 selected. The 'Dual ESI (Seg)' parameters are: Gas Temp 300 °C, Drying Gas 11 l/min, Nebulizer 50 psig, and a second set of values (300 °C, 3.0 l/min, 15 psig). The 'Dual ESI (Expt)' parameters are: VCap 3500 V, Capillary 0.044 uA, and Chamber 2.67 uA. The 'MS TOF (Expt)' parameters are: Fragmentor 135 V, Skimmer 65 V, OCT 1 RF Vpp 750 V, and Collision Energy 20 V (circled in red). The 'Cycle Time' is set to 1 s.

Time (min)	Expt
0	1
0.5	2
	3
	4

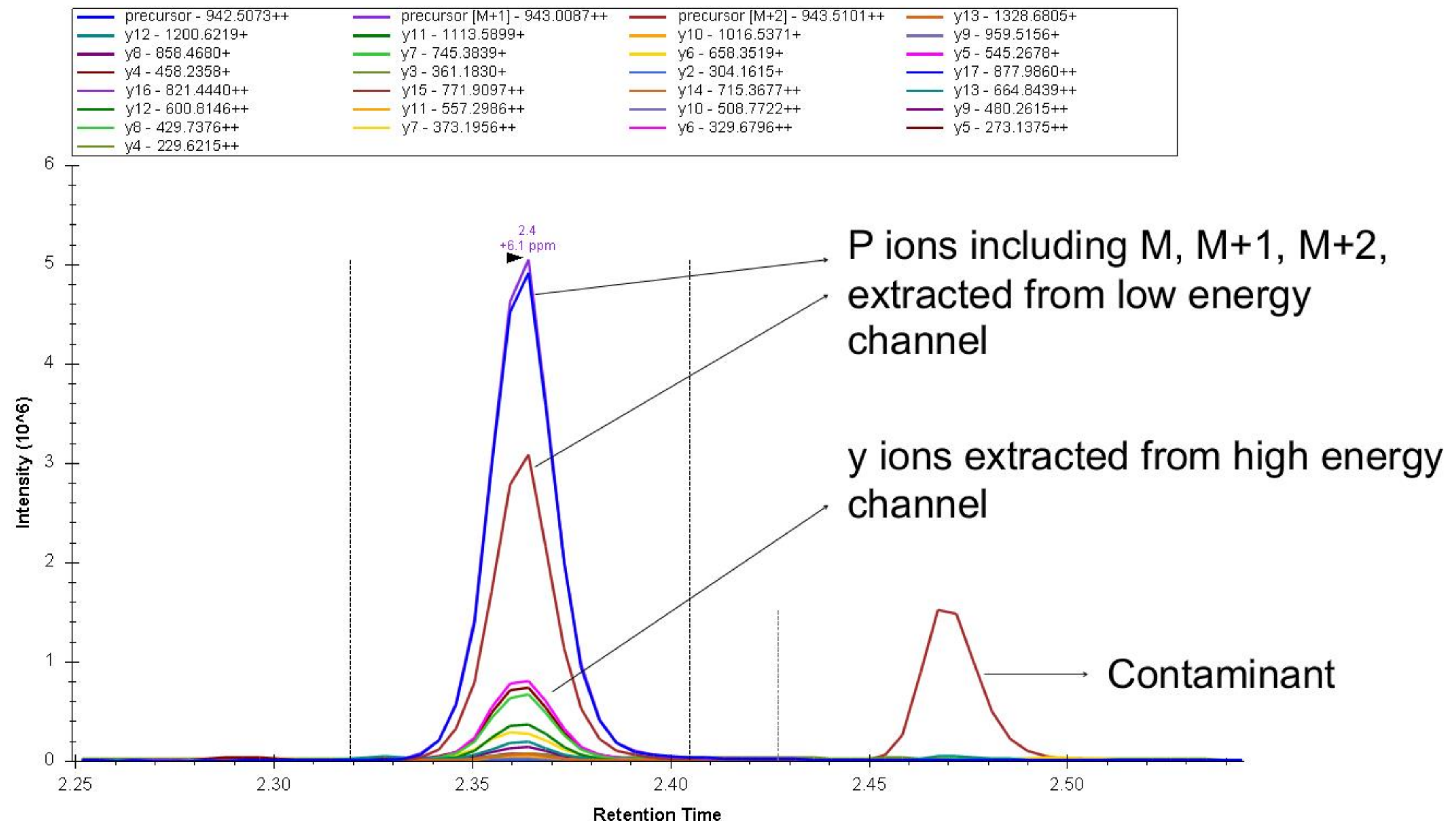
All Ions MS/MS on Agilent QTOF



All Ions → Skyline Workflow

Step 1: Show p & y Ions for Peptide Confirmation

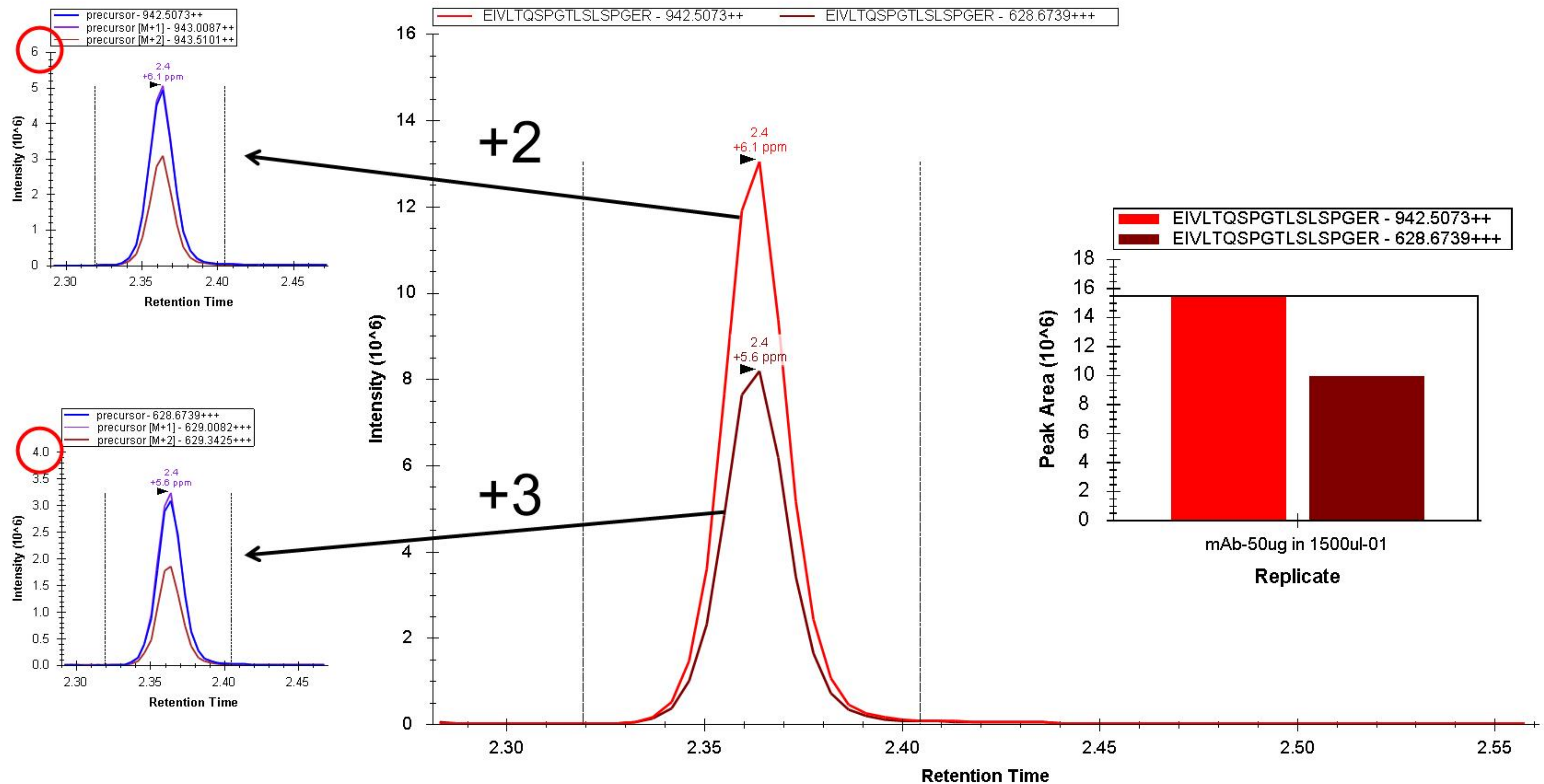
- Precursors (**p ions**) from low energy channel and products (**y ions**) from high energy channel are automatically extracted and plotted together to easily confirm the identity of the peptides.



All Ions → Skyline workflow

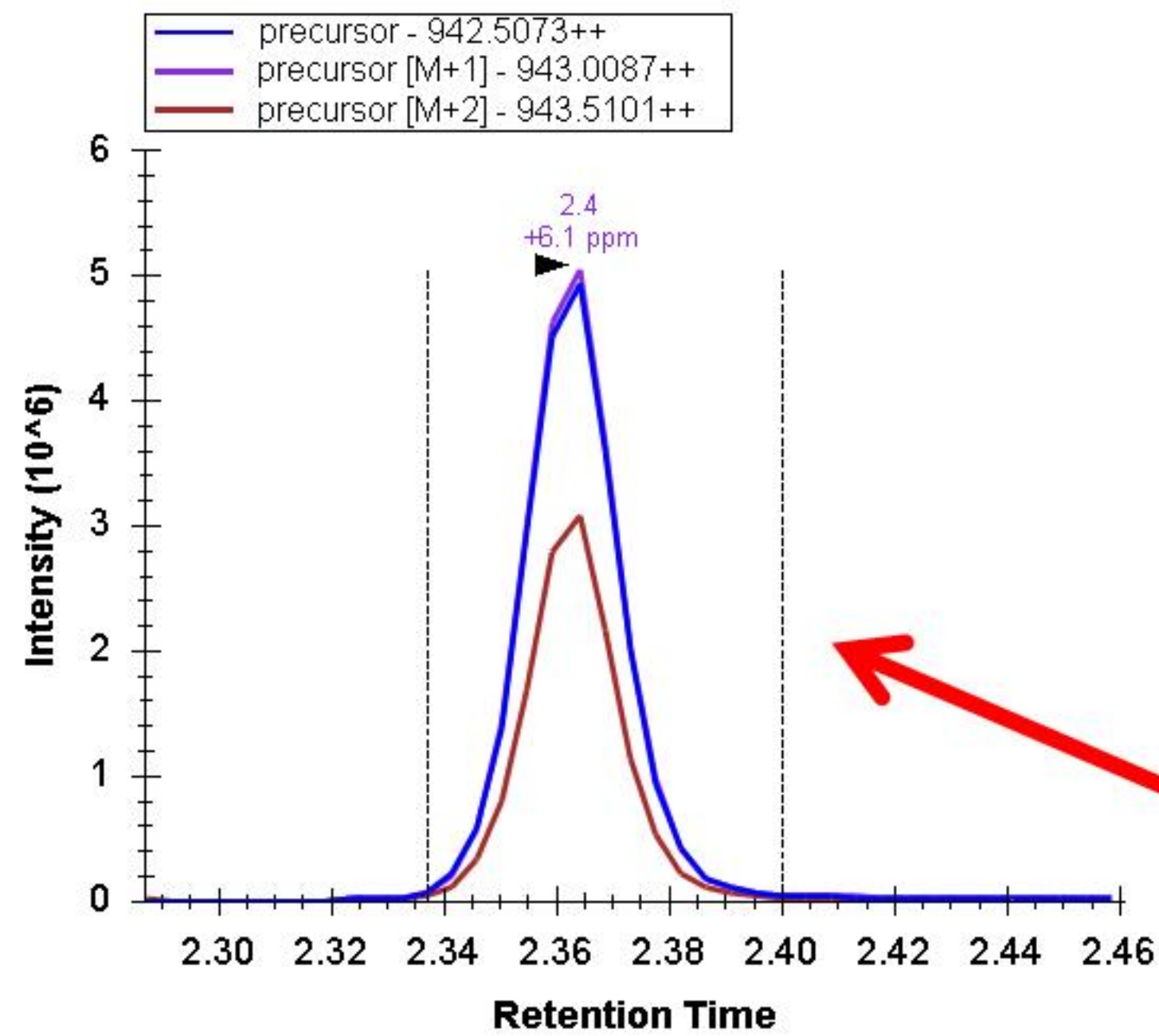
Step 2: Show only p ions to Select Best Charge State

Best charge state for each peptide is easily determined by comparing p ions only

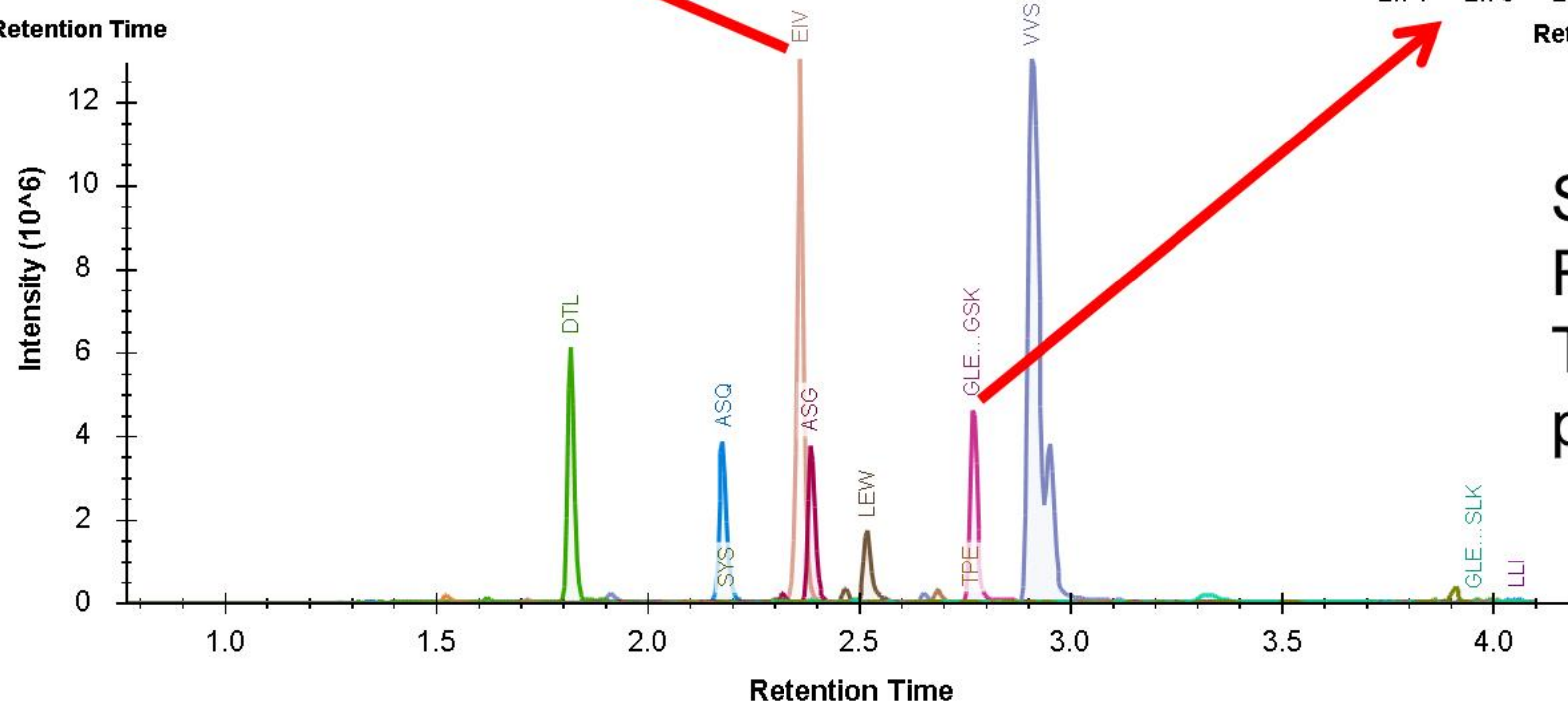
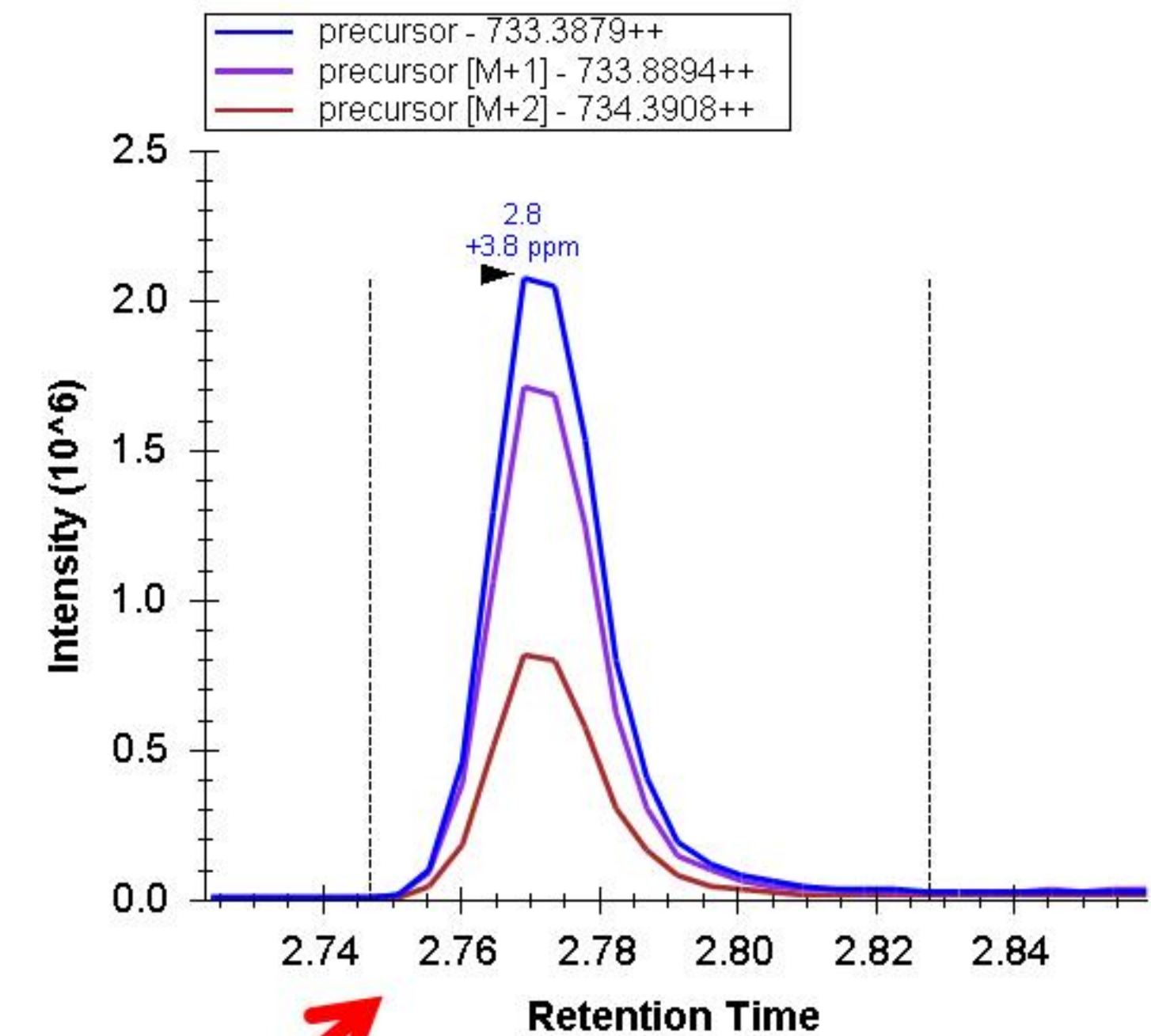


All Ions → Skyline workflow

Step 3: Show only p ions to Compare Peptide Abundance



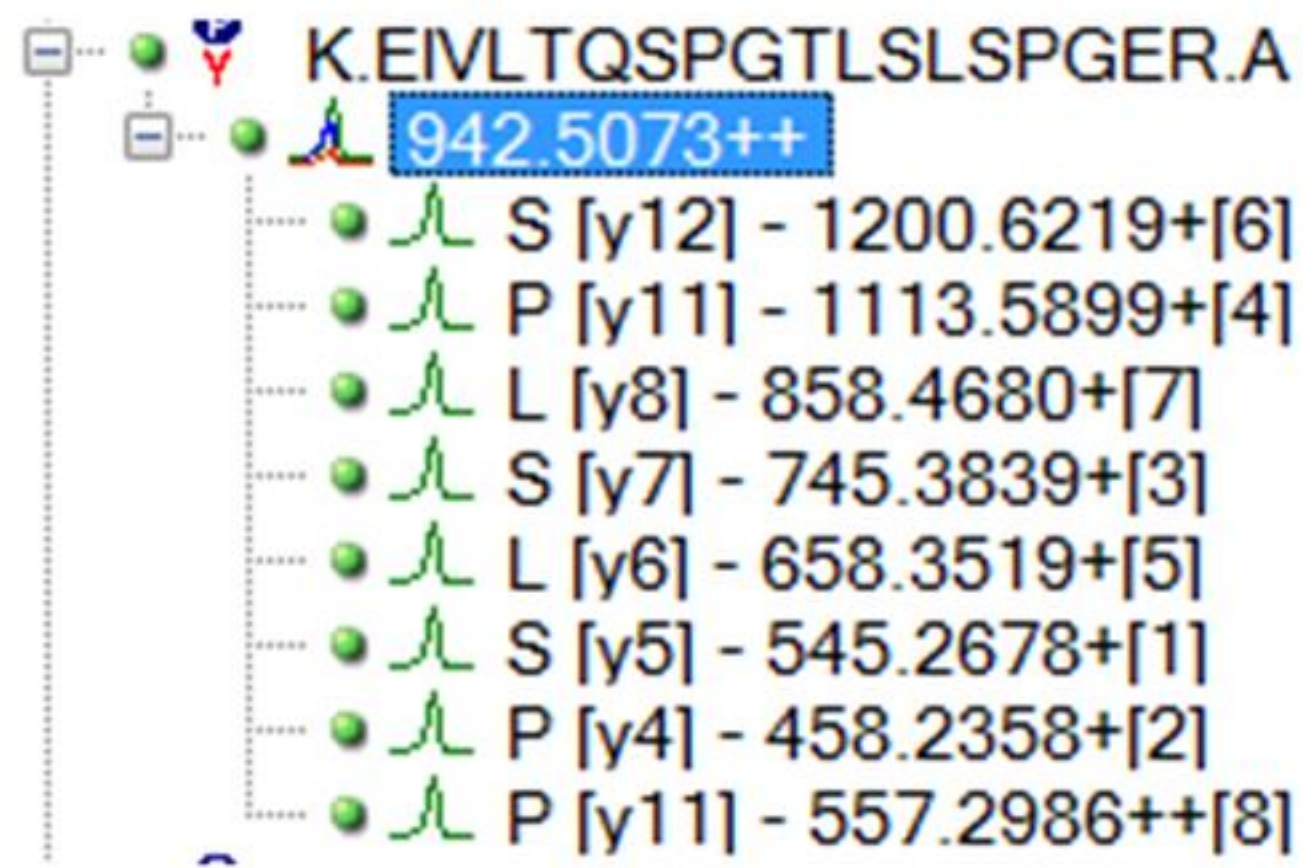
Peptides with the highest signal are easily determined by comparing p ions from the best charge state of all peptides.



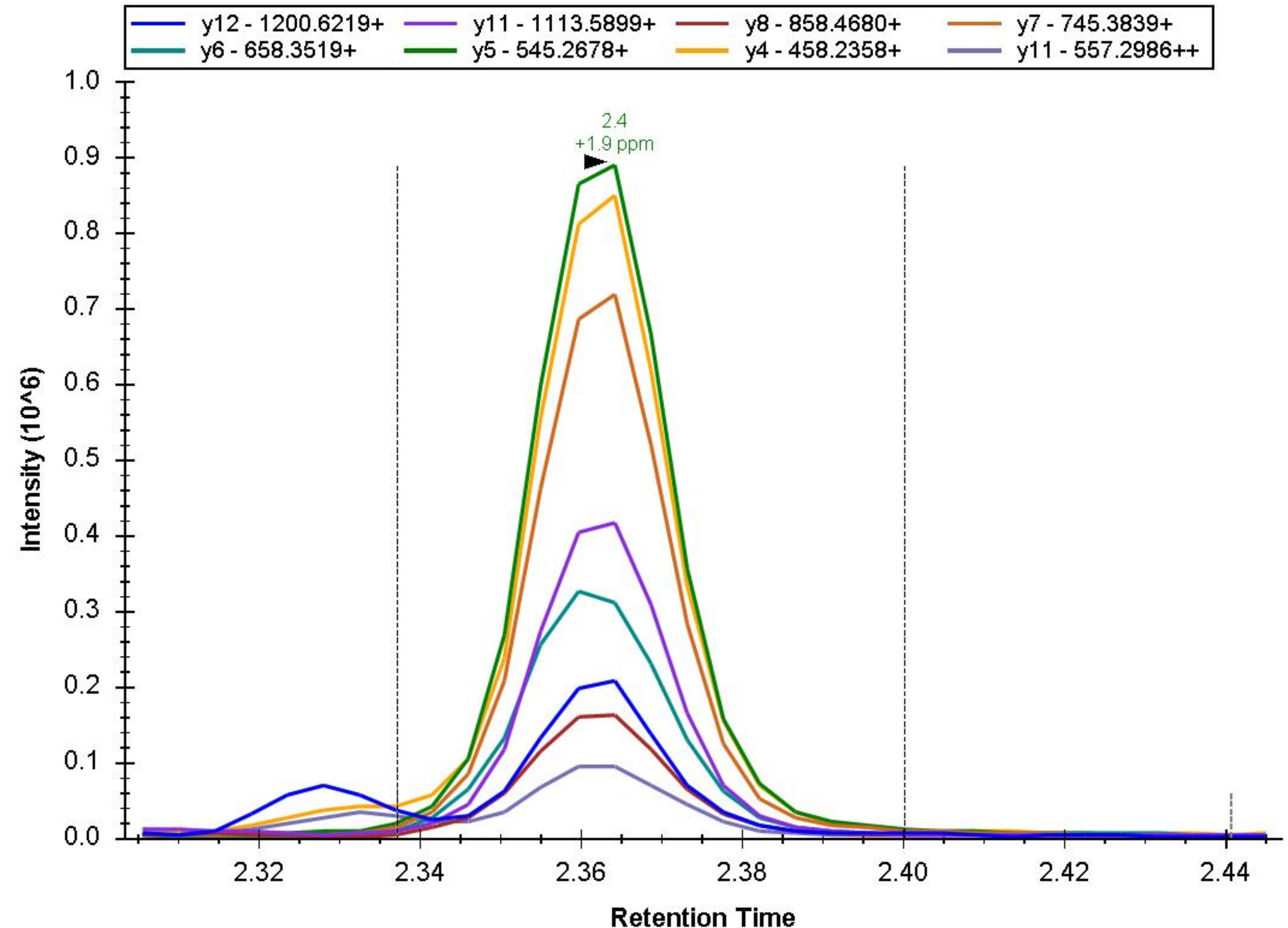
Skyline peptide
Rank Refine
Tool picks top n
peptides

All Ions → Skyline workflow

Step 4: Show Only y Ions to Select Top MRMs

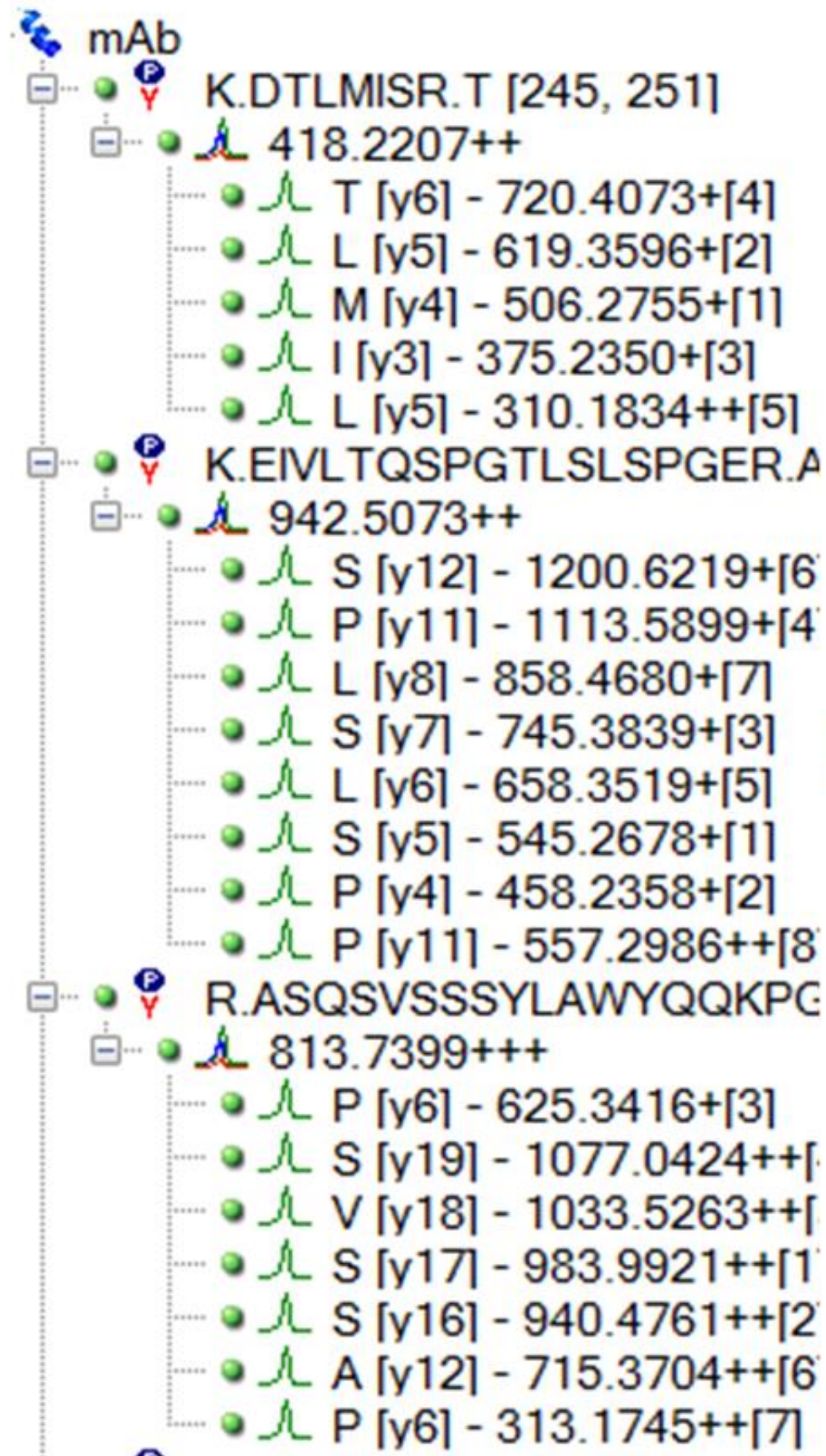


Skyline Transition
rank refine tool
picks top n MRMs



All Ions → Skyline workflow

Step 5: CE Optimization



Export Method

Instrument type: Agilent 6400 Series

Single method

One method per protein

Multiple methods

Ignore proteins

Max transitions per sample injection: 100

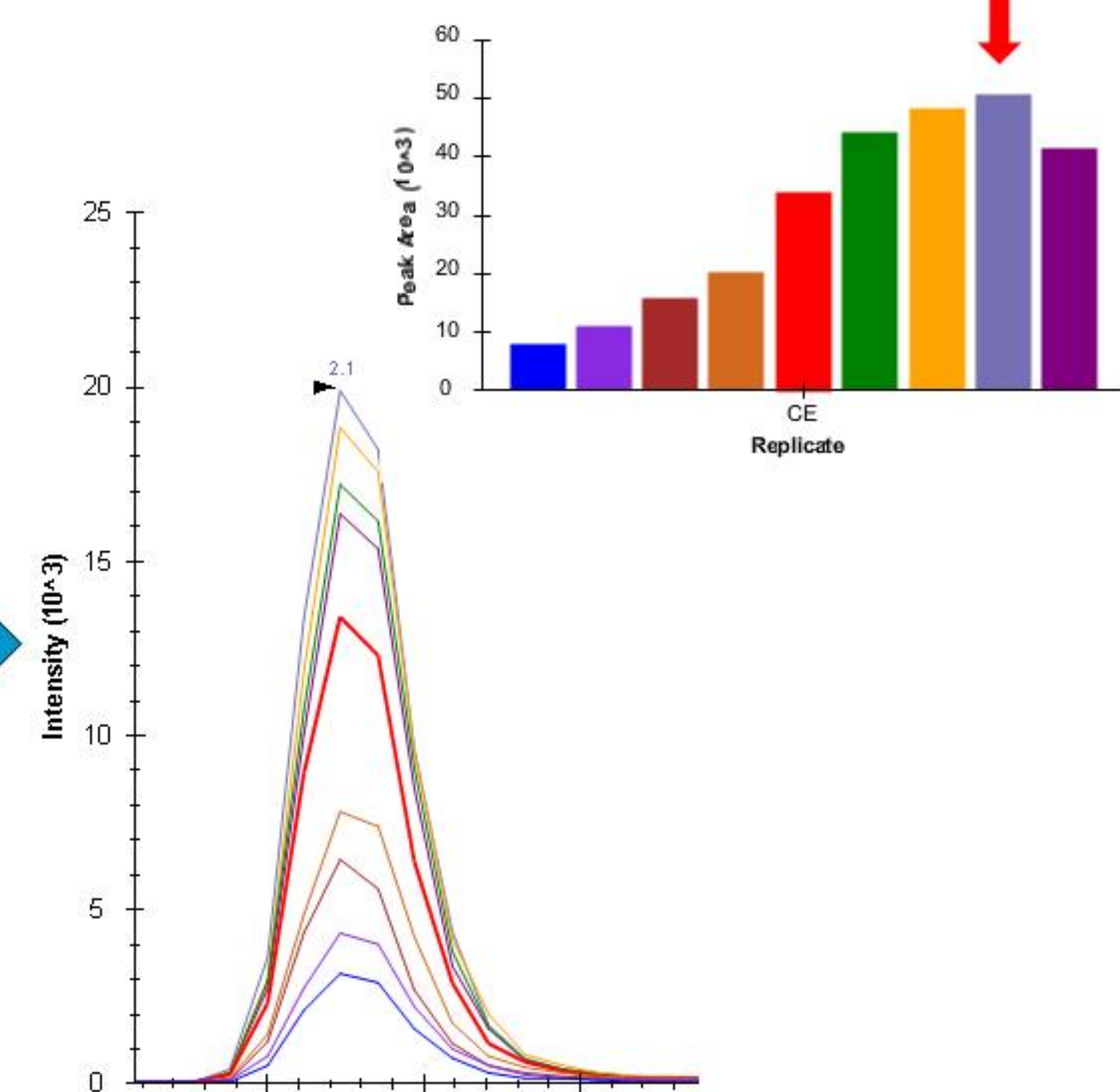
Methods: 2

Optimizing: Collision Energy

Method type: Standard

Dwell time (ms): 10

Template file: Browse...



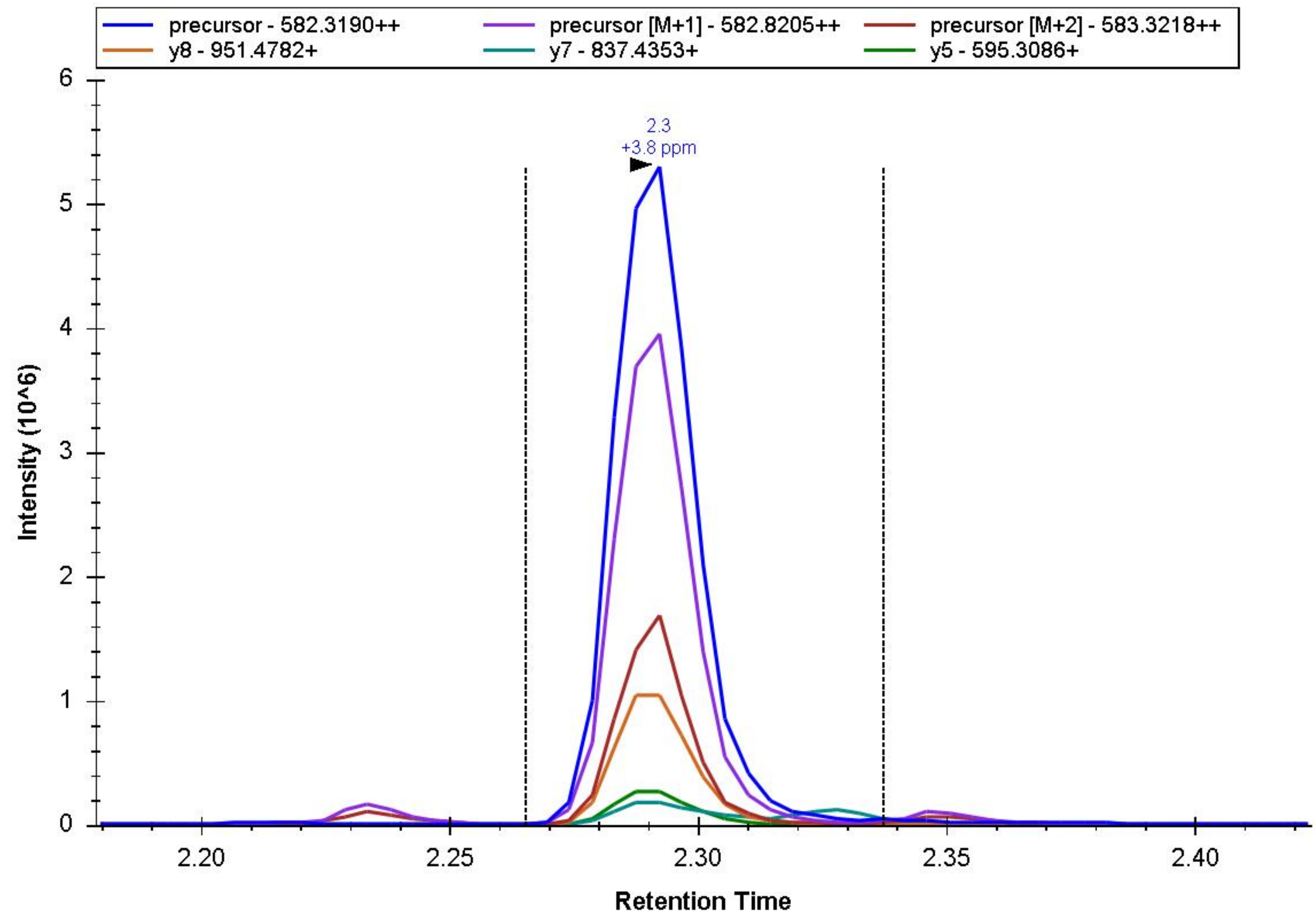
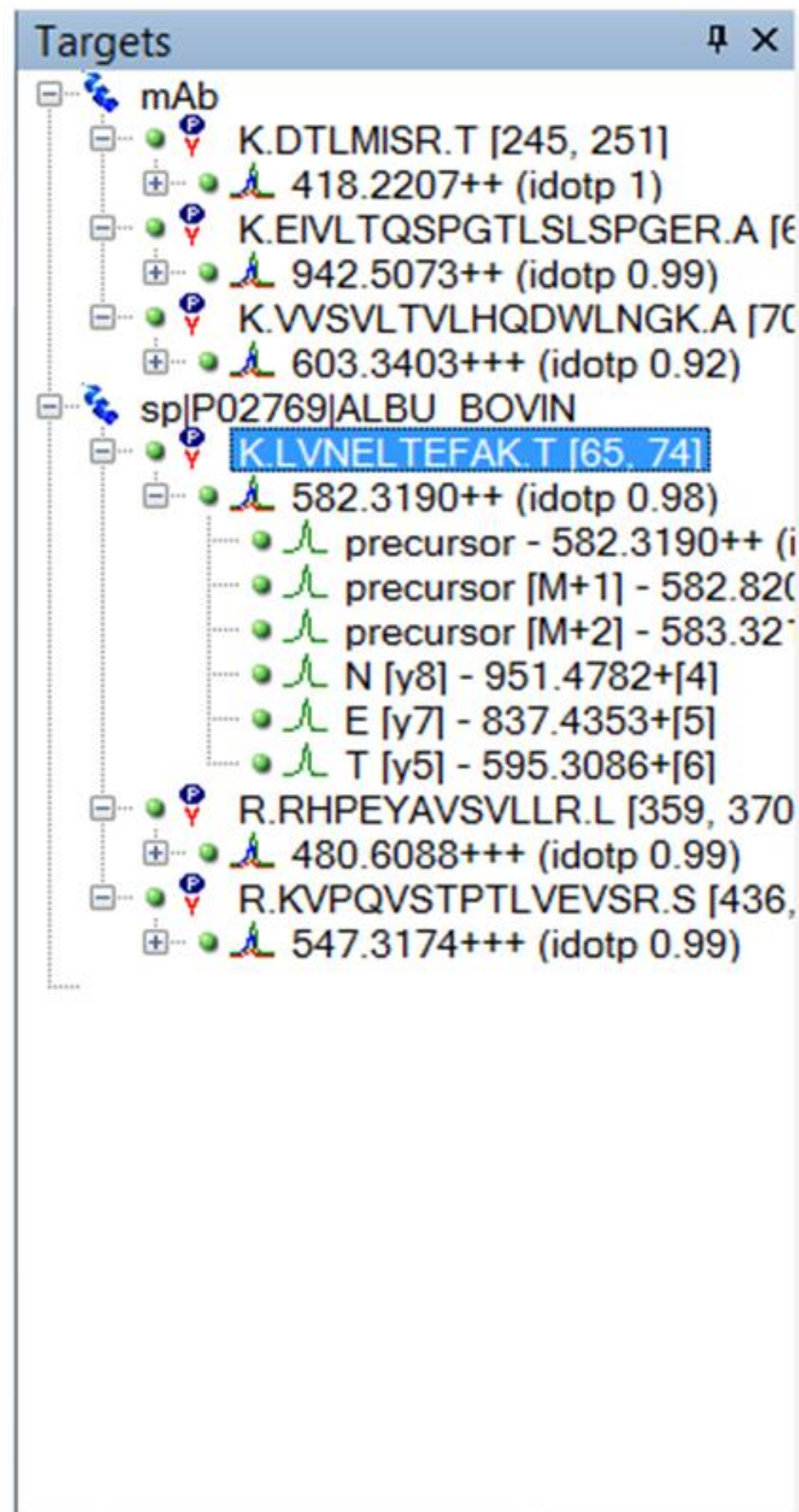
CE optimization process can be automated using new **Agilent Automation Tool** integrated into Skyline SW

Skyline Prediction Workflow vs. All Ions-Skyline Workflow

mAb Tryptic Digest	Skyline Prediction Workflow	All Ions – Skyline Workflow
Number of peptides for CE optimization	40	3 (Selected by All Ions run)
Injections required for CE optimization	40 (One inj. for one peptide)	3
Total time required for MRM method development	$40 \times 10 = 400$ min	$(3+1) \times 10 = 40$ min (1 is for All Ions run)

Save up to 90% of time and 90% of samples needed for CE optimization

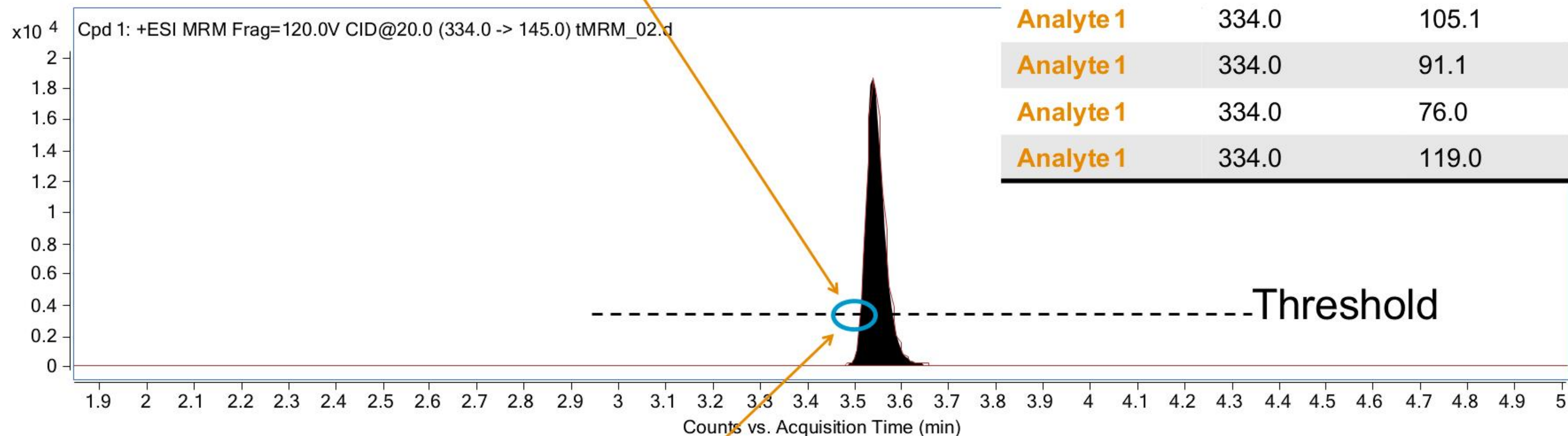
Method Development for Multiple Proteins



- Method development for multiple proteins can be done from the same All Ions result, simultaneously or at a later time.
- Import a different protein sequence and load the same All Ions data. Same procedure followed.

Triggered MRM (tMRM) Analysis

Secondary MRM
Transitions are "Triggered"



Triggered cycle (above threshold)

Analyte 1	334.0	145.0
Analyte 1	334.0	117.0
Analyte 1	334.0	132.1
Analyte 1	334.0	105.1
Analyte 1	334.0	91.1
Analyte 1	334.0	76.0
Analyte 1	334.0	119.0

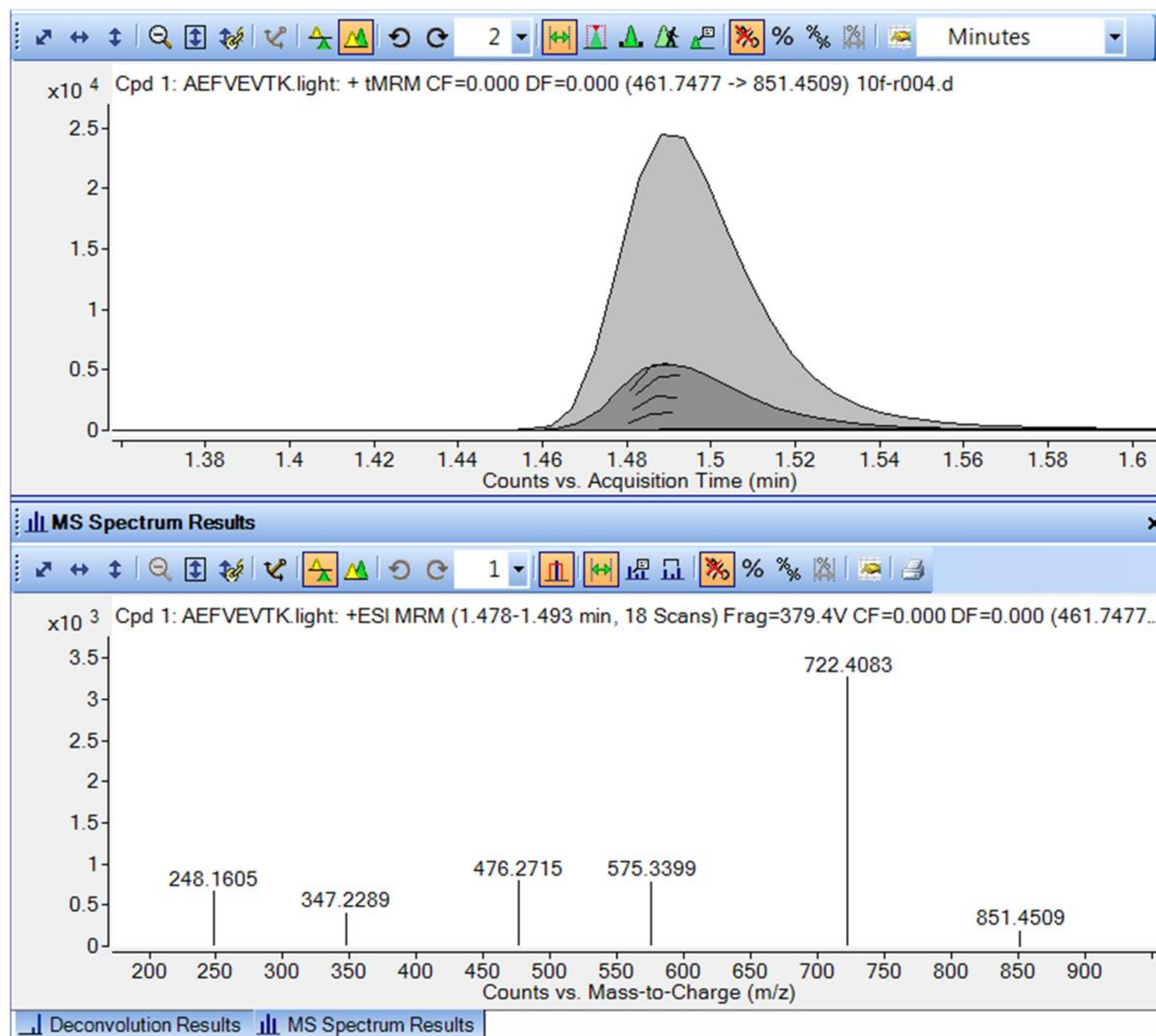
Primary cycle (below threshold)

Analyte 1	334.0	145.0
Analyte 1	334.0	117.0

Less cycle time comparing to product scan
Secondary transitions only collected for compounds found
to be present



tMRM Example



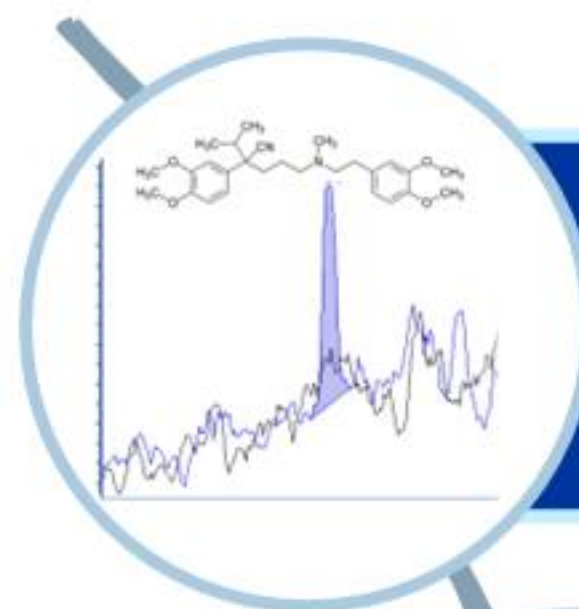
All Ions-Skyline Workflow Summary

With new All Ions – Skyline method development workflow:

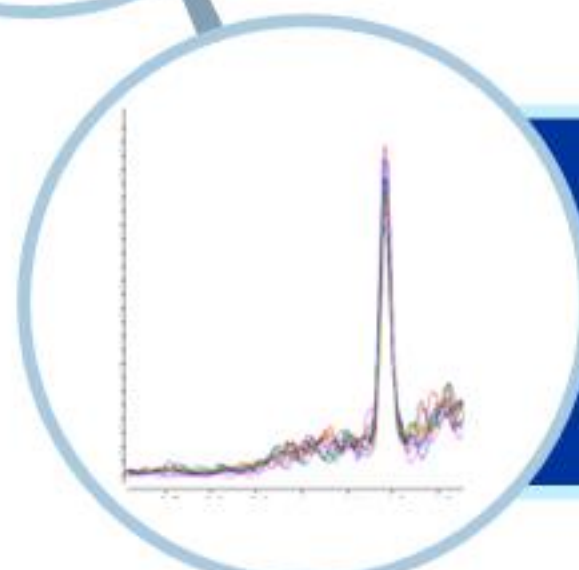
- MRM method development is much faster and simpler
- Agilent automation tool in Skyline SW automatically optimizes CE, creates and exports final QQQ method and runs the worklist
- Database search or spectral library is not needed
- Can be used for single protein or multiple proteins method development, as well as revisit for different proteins without more sample runs



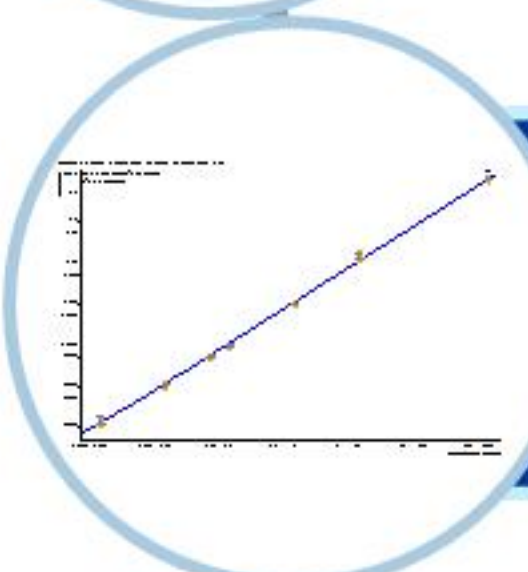
6495 QQQ LC/MS - Premium Performance



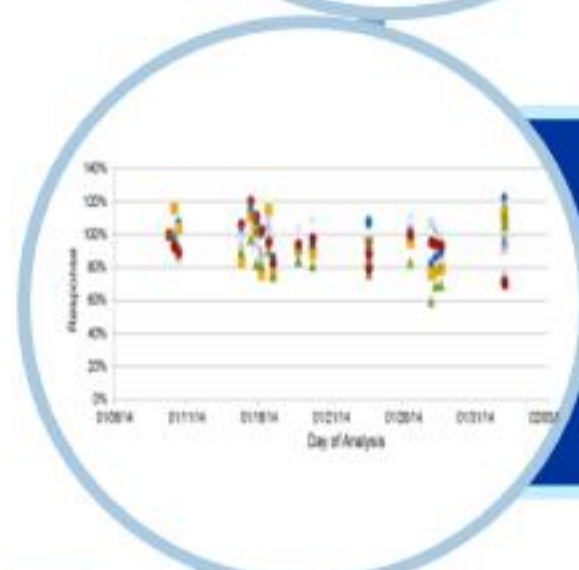
Improved sensitivity (IDL / MDL) – Average 3x in S/N specifications and applications



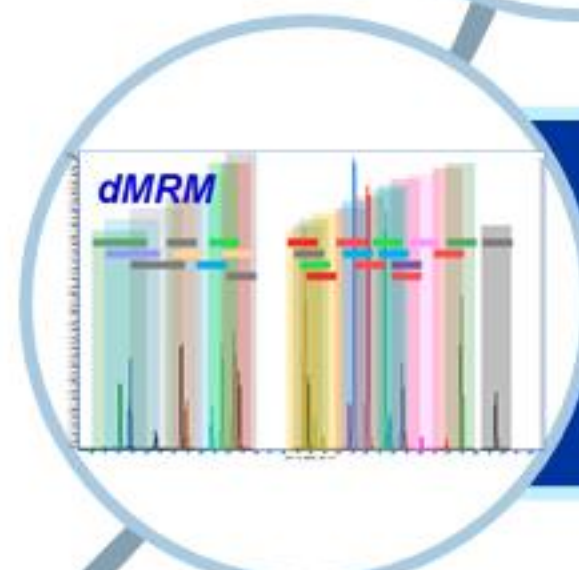
Improved precision and excellent accuracy at the lowest levels – 4 to 5x in IDL spec



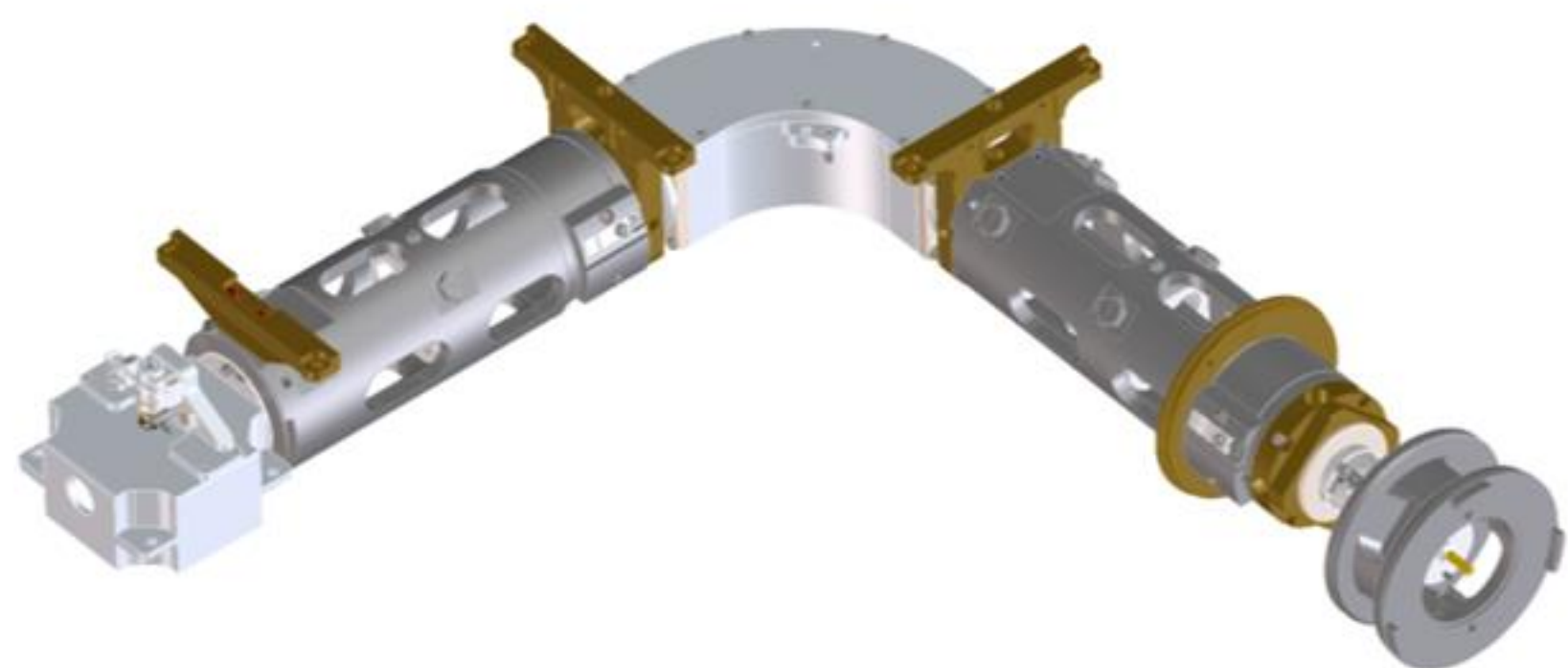
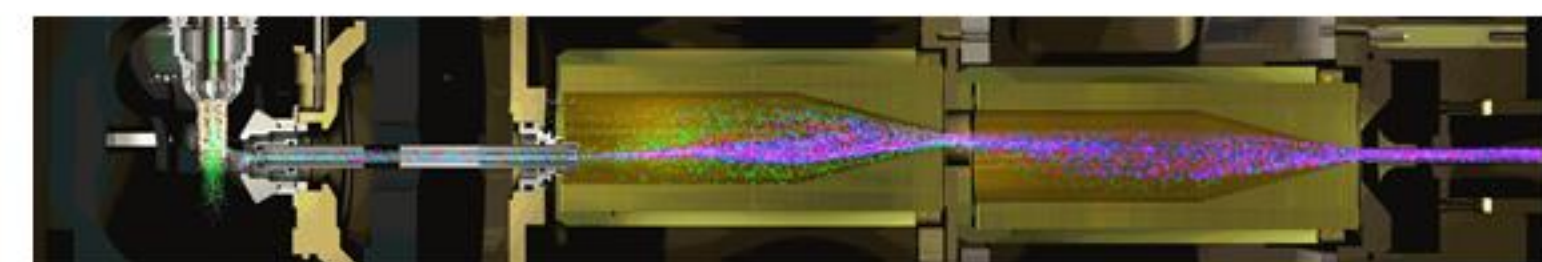
Proven 6 orders of linear dynamic range



Proven robustness in complex matrix – Food matrix and biological matrix (plasma)

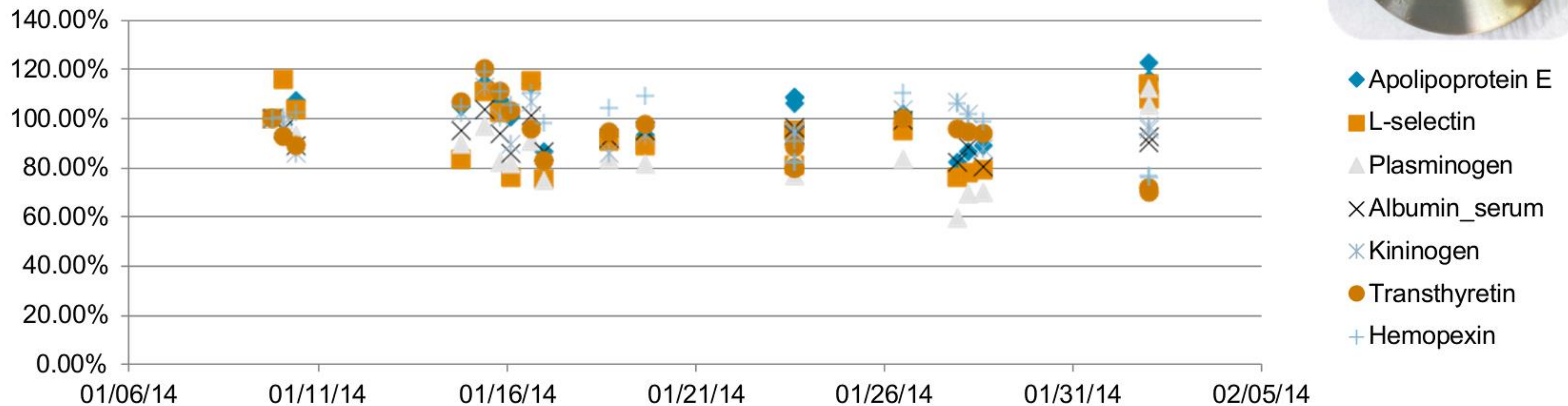


Improved mass range, fast scan speed and MRM acquisition rate



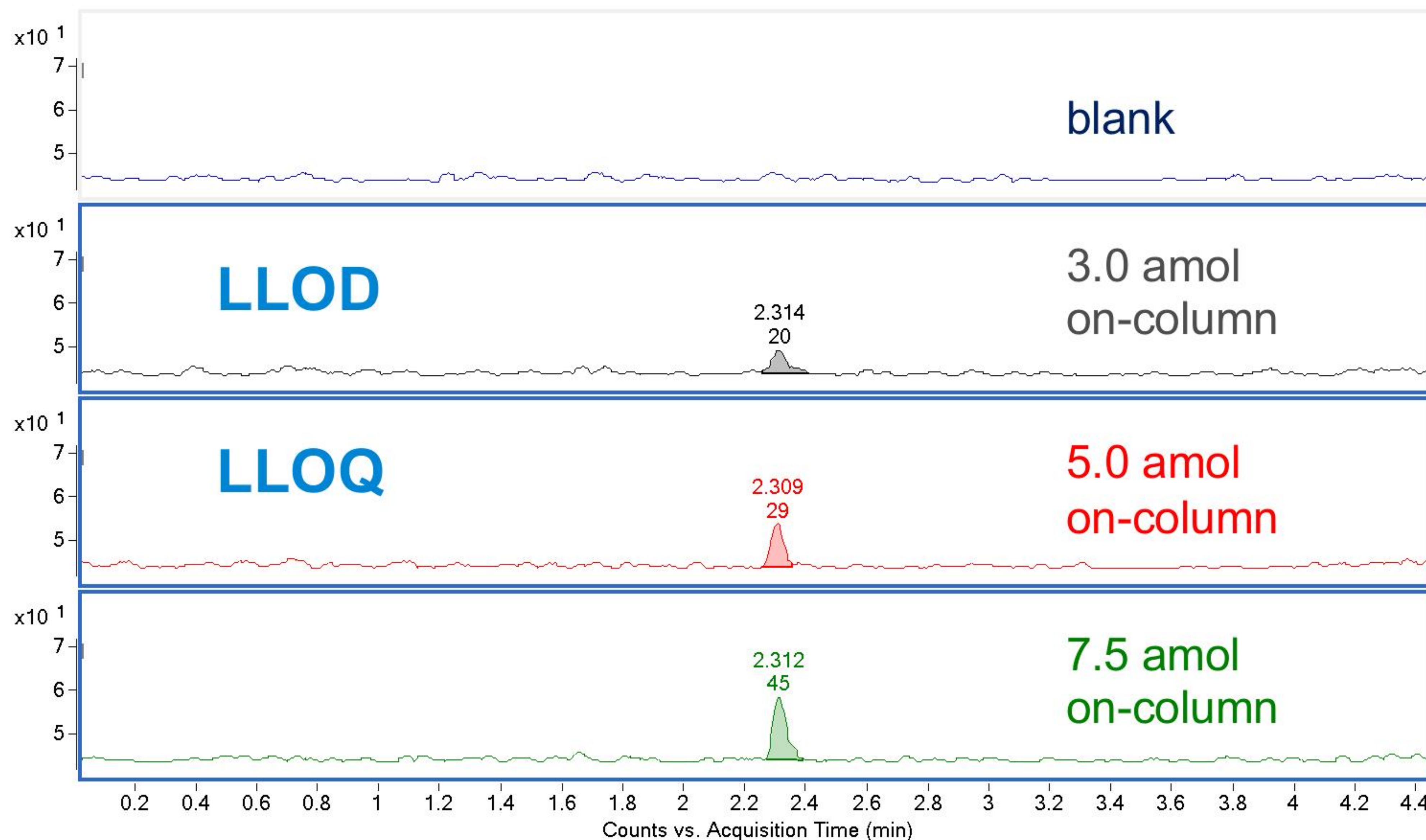
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Proven System Robustness in Complex Matrix: Protein Quantitation in Plasma



- Selected peptides from 42 peptides in the QC sample – normalized to Day 1 response
- **Peptide QC samples** analyzed daily after every ~25 plasma digest injections
- No significant signal degradation observed after **853 injections** of **40 µg plasma digest per injection** and **3.5 weeks of continuous operation**
- Response %RSD: 6 - 15

Peptide Quantitation: Outstanding Sensitivity with Standard Flow Chromatography

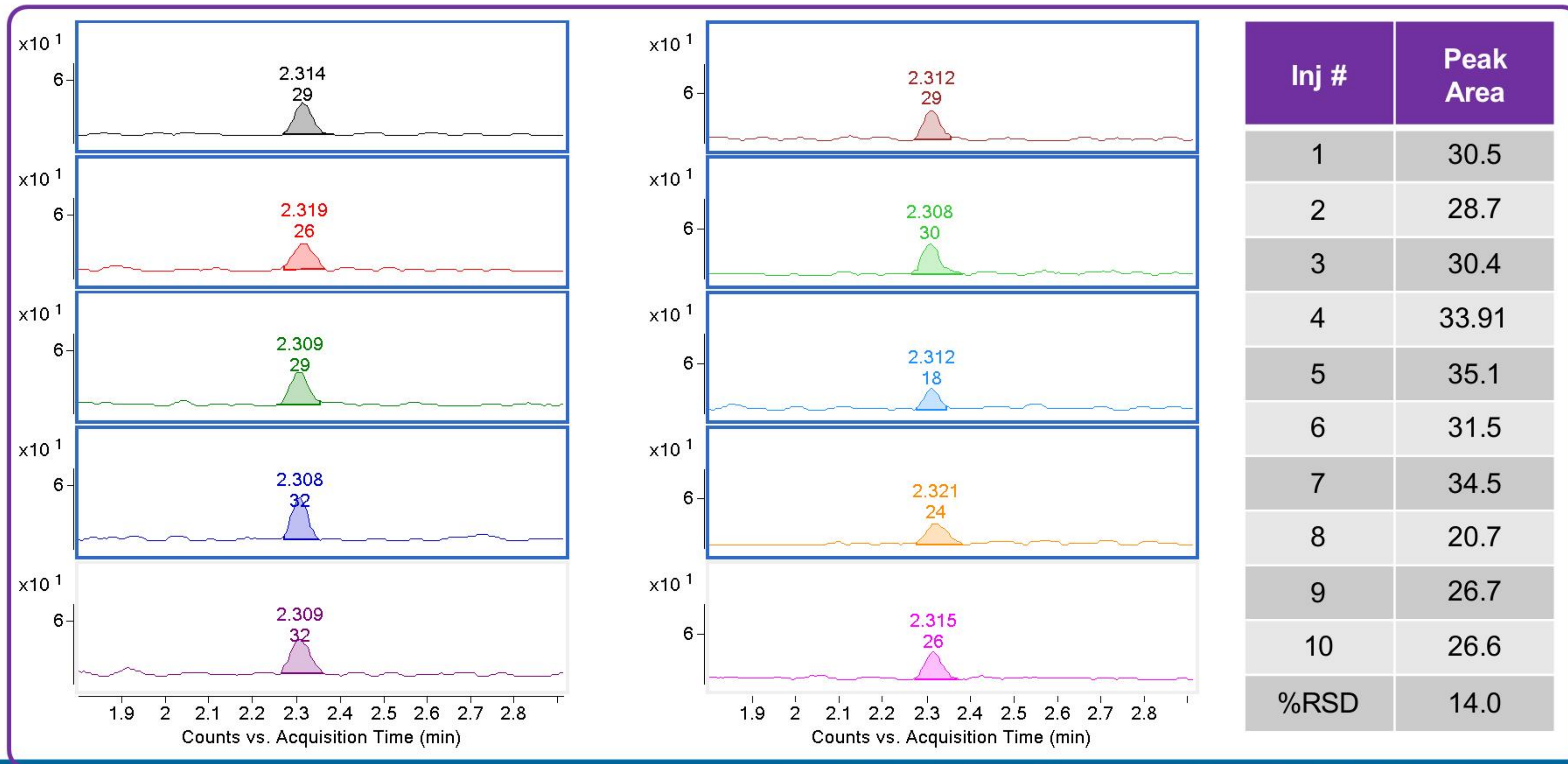


Instrumentation: Agilent JetStream + 6495 QQQ + 1290 UHPLC (2.1mm column, 0.6ml/min)
Sample: synthetic peptide standard (LVNEVTEFAK) spiked into enolase tryptic digest
Injection volume: 1 μ L

LVNEVTEFAK: Excellent Precision and Low Amol IDL

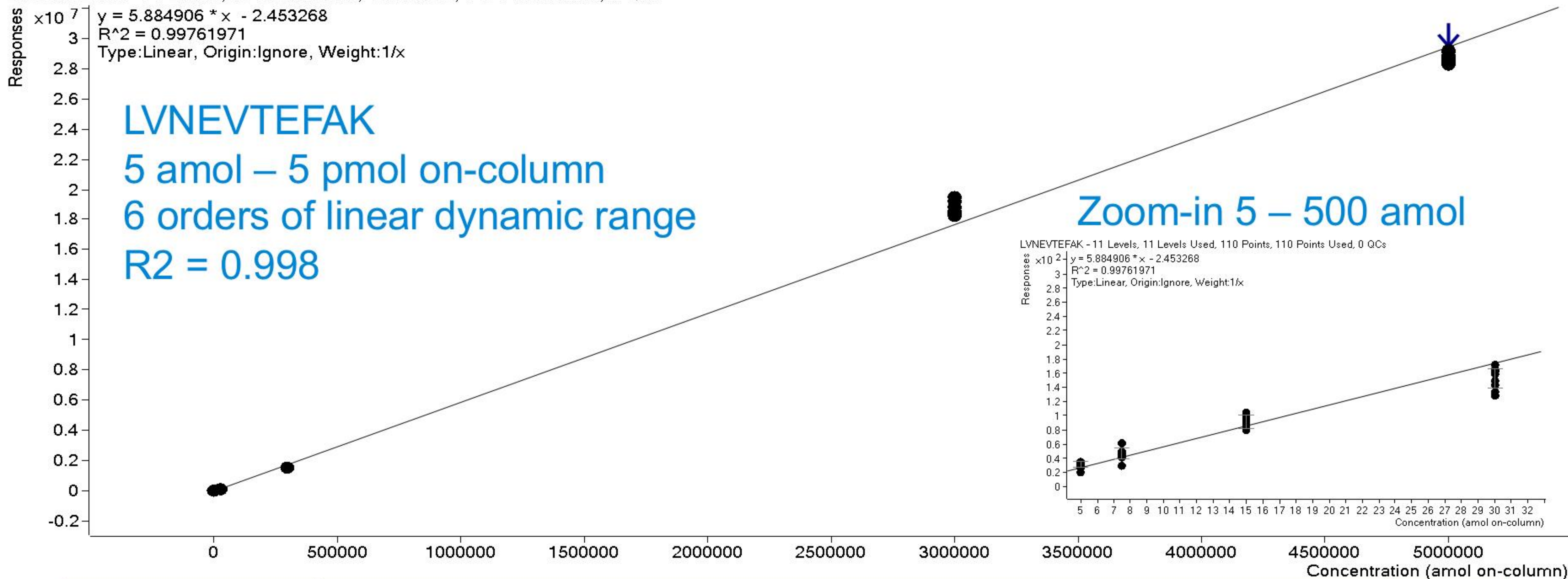
Amount measured	Replicates	%RSD	t (99%)	IDL
5.0 amol (LLOQ)	n = 10 injections	14.0	2.821	2.0 amol

$$\text{IDL} = t \times (\% \text{RSD} / 100) \times \text{Amount} = 2.821 \times (14.0 / 100) \times 5.0 \text{ amol} = 2.0 \text{ amol}$$



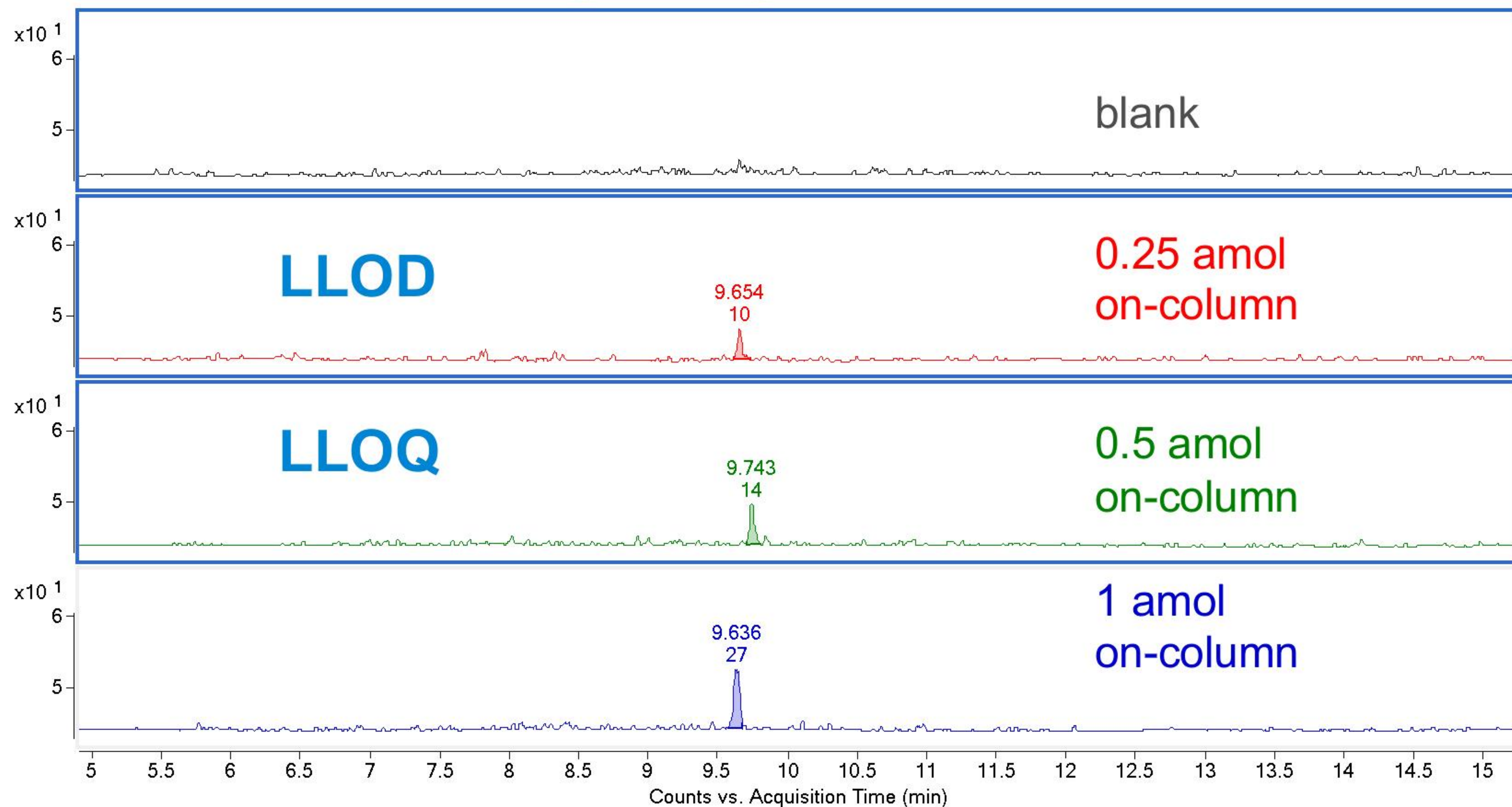
Peptide Quantitation – 6 Orders of Linear Dynamic Range

LVNEVTEFAK - 11 Levels, 11 Levels Used, 110 Points, 110 Points Used, 0 QCs



LVNEVTEFAK	Calibration Standards (amount on-column; 1 μ L injected)									
	5.0 amol	7.5 amol	15 amol	30 amol	300 amol	3 fmol	30 fmol	300 fmol	3 pmol	5 pmol
%Accuracy	109.8	108.7	105.0	87.1	85.2	81.4	86.4	87.4	105.6	97.5
Reproducibility (%RSD, n=10)	14.0	16.0	9.4	9.0	1.6	1.2	0.6	0.7	2.1	1.0
RT (%RSD, n=100)	0.12									

Peptide Quantitation: Ultimate Sensitivity with Nanoflow Chromatography



Instrumentation: 1260 HPLC-Chip/MS + 6495 QQQ

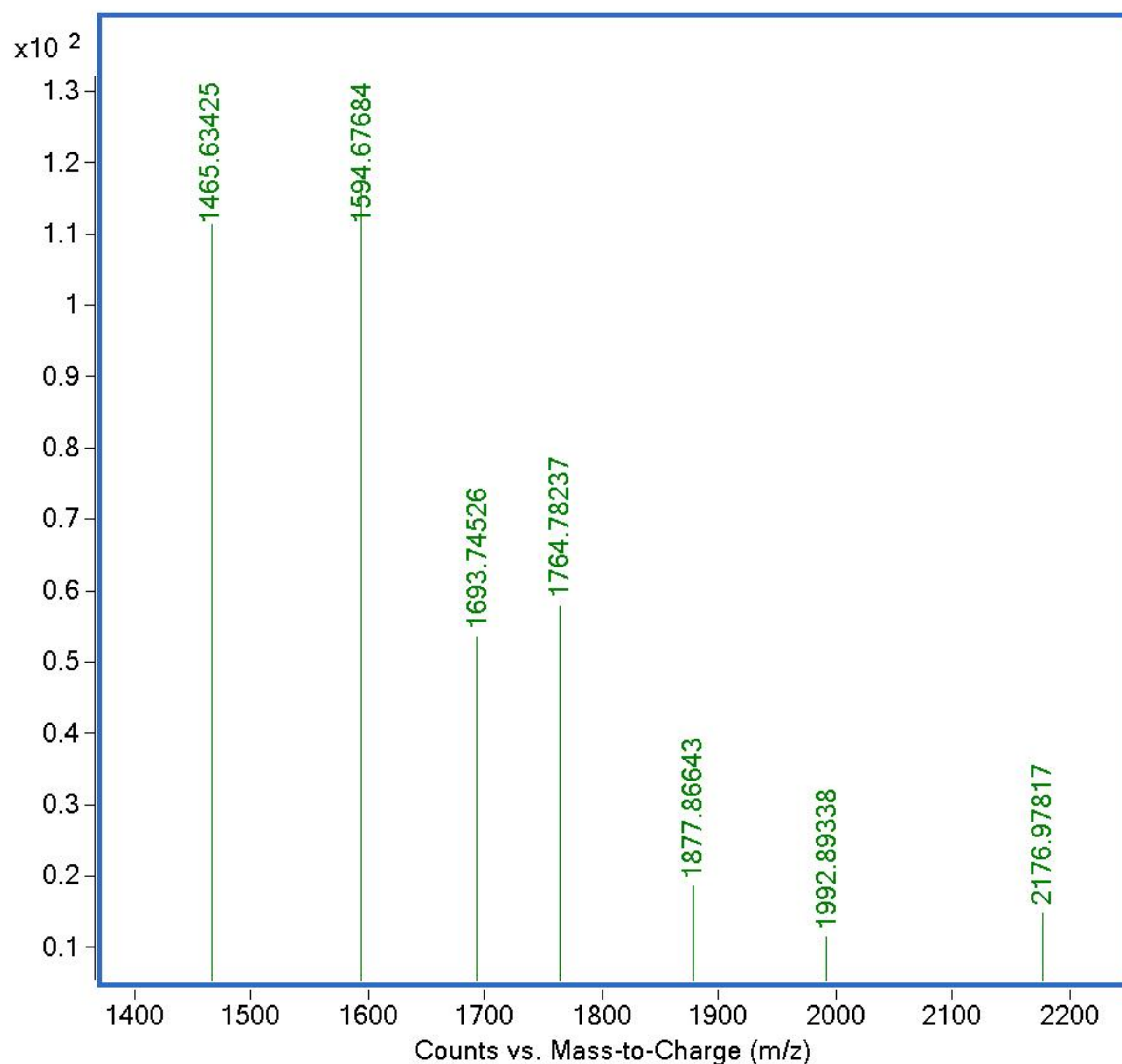
Sample: synthetic peptide standard (LVNEVTEFAK) spiked into enolase tryptic digest

Injection volume: 1 μ L

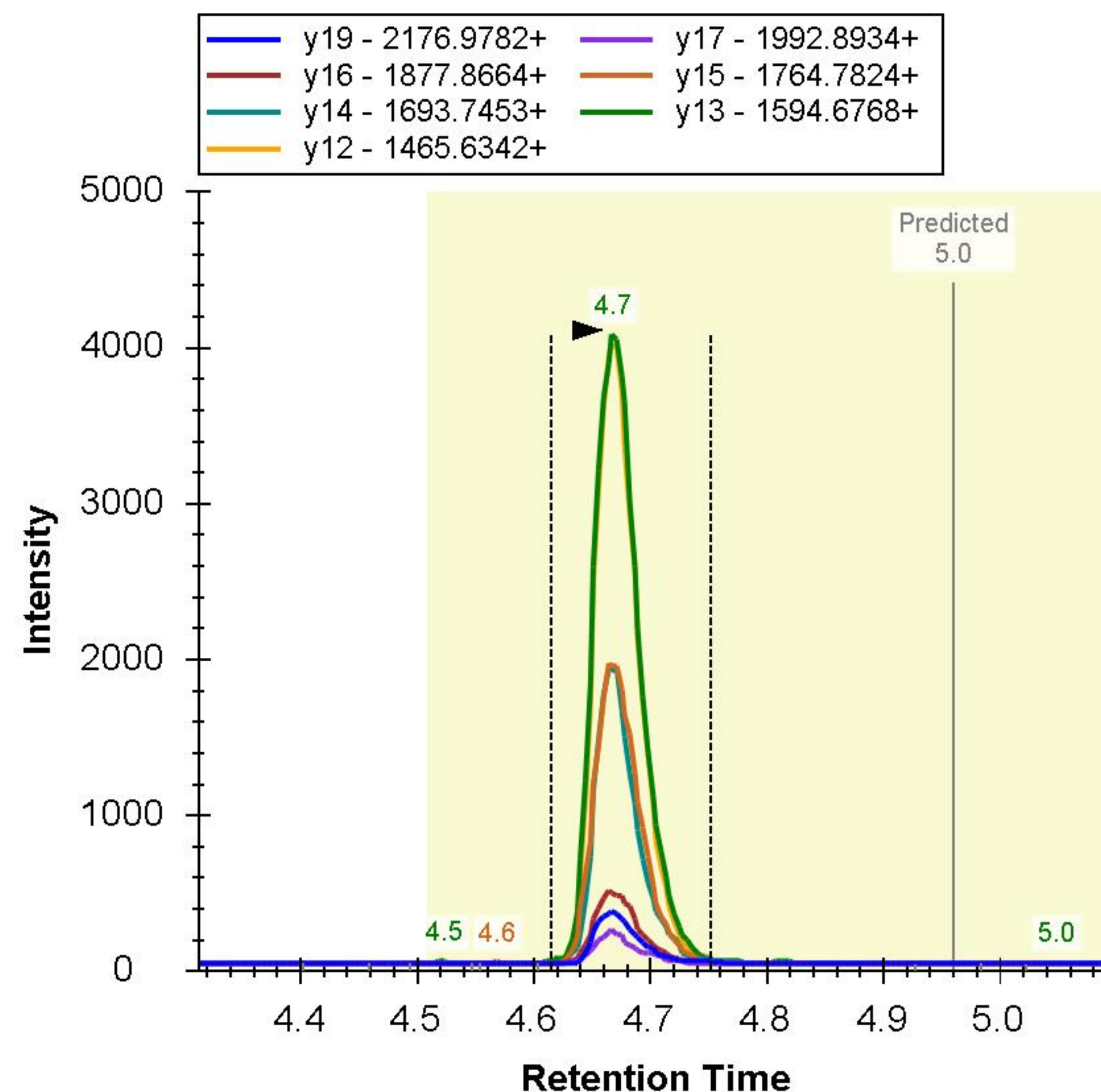


Increased Mass Range Useful for Peptides

MRM Spectrum



Skyline Overlay of MRM Chromatograms



MRM Spectrum for IgG Peptide: GFYPSDIAVEWESNGQPENNYK

Summary

- The Agilent 6495 Triple Quadrupole LC/MS achieves low attomole sensitivity for peptides.
- The calibration curves show excellent linearity with six orders of dynamic range with great accuracy and reproducibility.
- With a complex plasma digest matrix, the 6495 QQQ showed excellent response and retention time reproducibility over an extended period (Total of 34mg of plasma digest injected over 3.5 weeks!)
- Improved performance in peptide quantitation can be achieved by the increased precursor ion transmission, enhanced detection efficiency (HED voltage up to -20kV) and extended m/z range of the 6495 QQQ



THANK YOU !

